Natural Products

DOI: 10.1002/ange.201107730

Total Syntheses of Hopeanol and Hopeahainol A Empowered by a Chiral Brønsted Acid Induced Pinacol Rearrangement**

Scott A. Snyder, Stephen B. Thomas, Agathe C. Mayer, and Steven P. Breazzano*

Although the stilbene resveratrol is simple in terms of its size and functional group array, it possesses high chemical reactivity, a property that enables its conversion into hundreds of architecturally diverse bioactive oligomeric natural products.[1–3] Among recent dimeric isolates, hopeanol and hopeahainol A (**1** and **2**, Scheme 1) are two of the most intriguing given their constrained, partially dearomatized bicyclic cores and potent activity in antitumor and acetylcholinesterase inhibition assays.[4] Indeed, these molecules have already been the subject of synthetic interest, with reports by Nicolaou et al. describing racemic and enantioselective syntheses of **1** and **2** in 15 linear steps.[5] Their route featured several cascade-based bond constructions[6] and the discovery that hopeahainol A (**2**) could be converted into hopeanol (**1**) upon treatment with base, an idea counter to the original biosynthetic proposal.[4b] Herein, we describe a distinct approach for the total synthesis of these natural products empowered by a unique, reagent-driven pinacol rearrangement and substrate-specific oxidation chemistry. Significantly, it has potential for scaleability as well as biogenetic implications.

Our retrosynthetic analysis is shown in the lower portion of Scheme 1, wherein our key disconnections were focused on rapidly constructing the seven-membered ring and attendant quaternary carbon center (C7b) found in both natural products, as best noted by a redrawing of **1** and **2**. Critical insights came following a change in the oxidation state of **2** to that of **3**, in that we anticipated that the all carbon-based quaternary center (C7b) could potentially arise from diol **5** through a pinacol rearrangement.[7] Although such events often possess modest selectivity as a result of ambiguity in the site of carbocation formation and/or migrating group, we hoped that the specific patterning of **5** could avoid such issues. Also, assuming that such a rearrangement could proceed with any stereoisomeric variant of **5**, then issues of diastereocontrol would not be a relevant concern, as all isomers of **3** should be able to be funneled to racemic **2** through oxidation chemistry. Issues in diastereocontrol occurred several times in the approach of Nicolaou et al. to this same ring system.[5] Additionally, we felt the complete route should be concise if the materials needed for this key rearrangement step could arise from ketone **6**, variants of which we synthesized previously through acid-induced cyclizations of alcohol **7**.[8] These materials have already enabled controlled syntheses of nearly 20 dimeric and higher-order natural products within the resveratrol class through several distinct, cascade-based constructions of diverse C–C and C–O bonds.[8,9] Finally, the route had two additional appealing elements. First, it is redox economic.[10] Second, it might possess biogenetic relevance given the structures of other seven-membered ring natural products. For example, if reactive compounds **8**–**10** were precursors[11] for natural products **11**–**14**[12] by proton cyclizations, then the same starting materials could lead to the C7b quaternary carbon center of **1** and **2** by initial oxidation (to generate **5** or a related congener) followed by acid treatment as likely needed to initiate pinacol rearrangement.[13]

We began our efforts by synthesizing diols of type **5**. As shown in Scheme 2, that goal was accomplished through a unique protocol starting from ketone **15** (prepared in five steps from commercial resveratrol in 48% overall yield, see Supporting Information),[14] a methyl ether protected version of **6** (compare with Scheme 1) redrawn with three-dimensional structure.[15] Following Corey–Chaykovsky epoxidation,[16] which afforded **16** with complete relative stereocontrol, subsequent dissolution in CH_2Cl_2 and stirring with AcOH at 25 °C generated what we believe to be the acetate-opened epoxide and/or an intermediate diol with inverted chirality at the C7b-position;[17] subsequent exposure to the Dess–Martin periodinane, followed by Grignard attack, afforded separable diols **19** and **20** in a 1:1.3 ratio.[18] Critically, the two ring-based chiral centers were formed with complete relative stereocontrol, an outcome that can be rationalized by the steric bulk of the remote aryl ring within **17**,[19] and one that proved essential to the success of the later sequence (see below). Worth noting is that other routes towards pinacol-type precursors were attempted, largely by trying to add nucleophiles to the ketone in **15**. However, none provided the expected materials with the exception of the Tebbe reagent; in this case, the resultant methylene group could not be functionalized further.

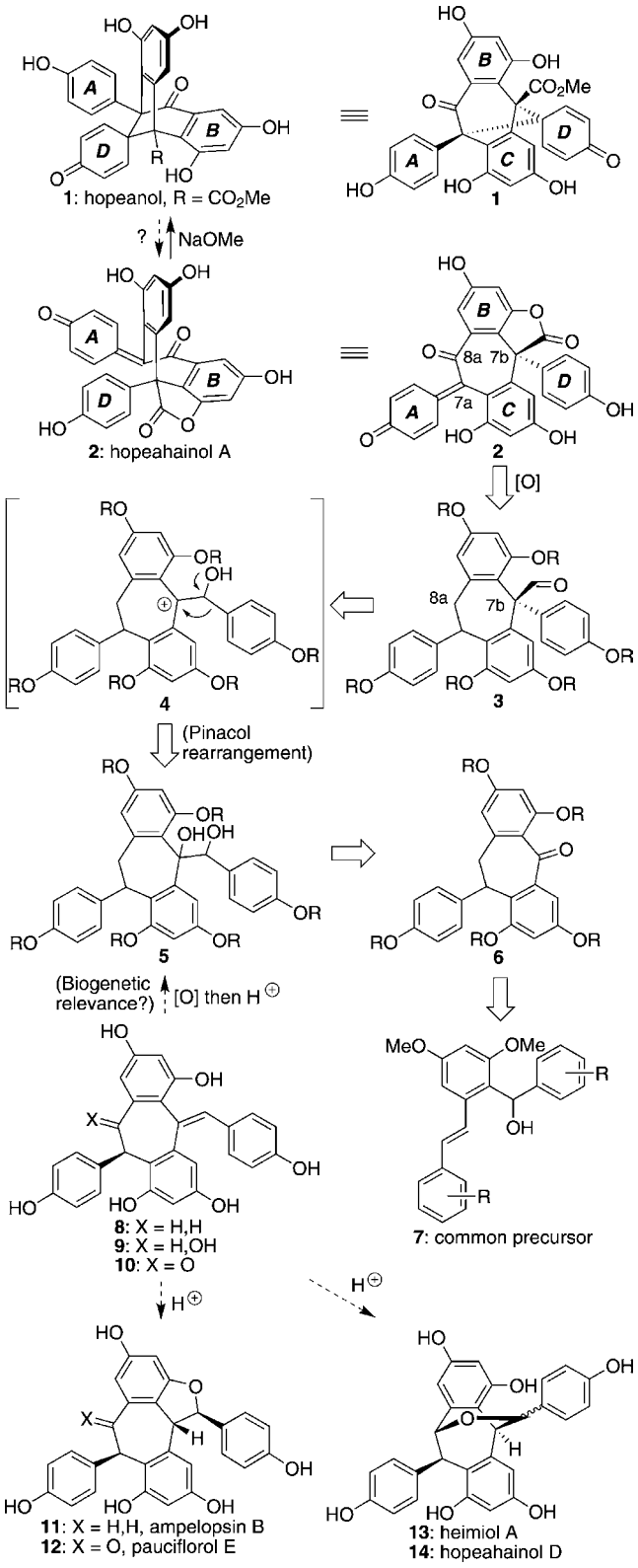
[*] Prof. Dr. S. A. Snyder, S. B. Thomas, Dr. A. C. Mayer,[+] S. P. Breazzano[+]
Dept. of Chemistry, Columbia University
Havemeyer Hall, 3000 Broadway, New York, NY 10027 (USA)
E-mail: sas2197@columbia.edu
Homepage: http://www.columbia.edu/cu/chemistry/groups/snyder/

[+] These authors contributed equally to initial studies.

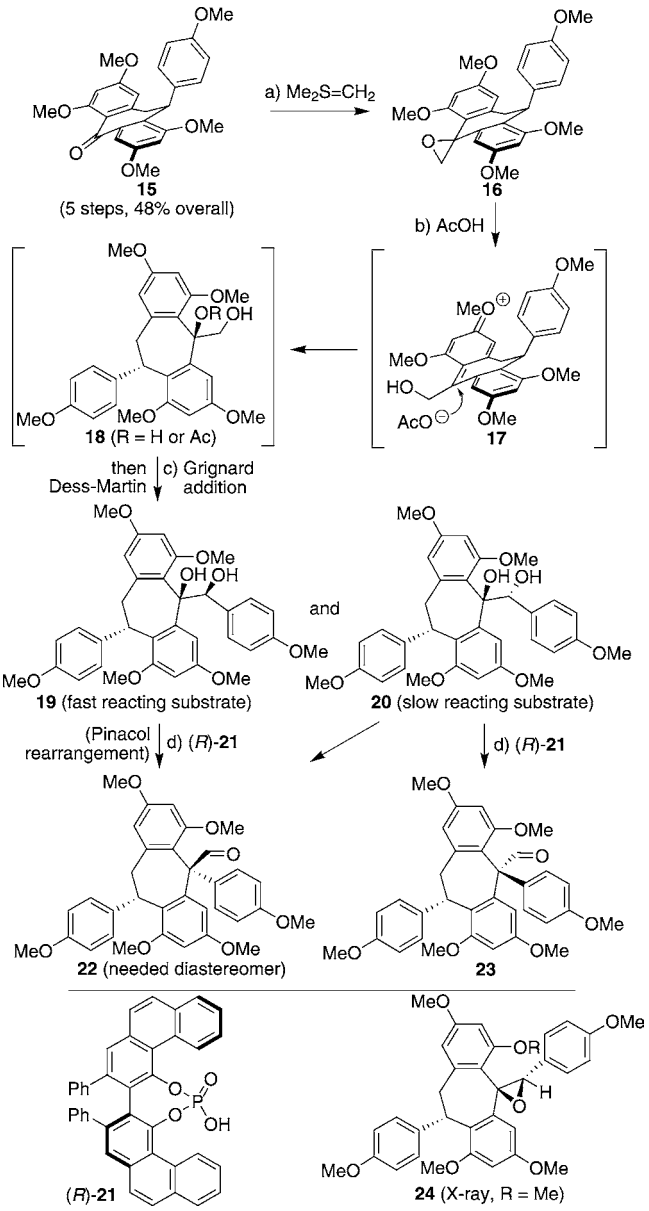
[**] We thank the NSF (CHE-0619638) for an X-ray diffractometer and Prof. Gerard Parkin, Wesley Sattler, Aaron Sattler, and Ashley Zuzek for performing the crystallographic analyses. We thank Adel ElSohly for NMR assistance and Jason Pflueger for early work. Financial support was provided by Columbia University, the National Institutes of Health (R01-GM84994), Bristol-Myers Squibb, Eli Lilly, the NSF (Predoctoral Fellowships to S.B.T. and S.P.B.), the DFG (Postdoctoral Fellowship to A.C.M.), and the Research Corporation for Science Advancement (Cottrell Scholar Award to S.A.S.).

Supporting information for this article is available on the WWW under http://dx.doi.org/10.1002/anie.201107730.

Nevertheless, with **19** and **20** in hand, explorations into the critical pinacol rearrangement could begin. Pleasingly, many protic and Lewis acids, such as *p*-toluenesulfonic acid (*p*-TsOH), pyridinium *p*-toluenesulfonate (PPTS), and trimethylsilyl trifluoromethanesulfonate (TMSOTf), could generate the desired quaternary carbon center of **22** and **23**, though there were some, such as benzoic acid, that did not. However, those that worked did so in low to moderate yield and with modest diastereocontrol, a critical issue as only **22** proved competent in later chemistry. Several side products were also observed in varying amounts, the most significant and consistent of which was epoxide **24**, a material whose structure was confirmed by X-ray analysis and which could not be converted into a pinacol rearranged product under any conditions.[20] A small subset of these initial results are

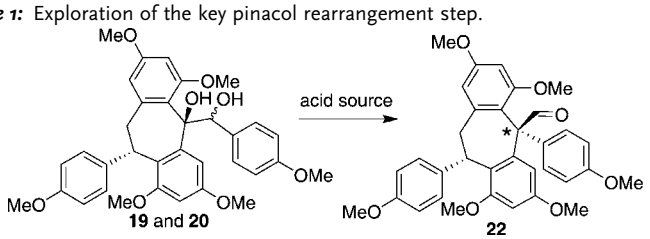


Scheme 1. Structures of hopeanol (**1**) and hopeahainol A (**2**) and retrosynthetic analysis based on a pinacol rearrangement as well as site-specific oxidations inspired by a potential biogenesis from **8**–**10**.



Scheme 2. Synthesis and pinacol rearrangement of **19** and **20**. a) Me_3SI (10 equiv), *n*BuLi (8.0 equiv), THF, 0 °C, 1 h. b) AcOH, CH_2Cl_2, 25 °C, 30 min; then Dess–Martin periodinane (1.2 equiv), 25 °C, 1 h, 45% over two steps. c) 4-OMePhMgBr (5.0 equiv), THF, 0→25 °C, 1.5 h, 87%, ca. 1.3:1 of **20**:**19**. d) (*R*)-**21** (1.0 equiv), $CHCl_3$, microwave, 100 °C, 1 h, 56% **22**/**23** as a >18:1 mix of diastereomers.

Table 1: Exploration of the key pinacol rearrangement step.



Ent	Acid	Equiv	Solvent	*T* [°C]	*t* [h]	Yield [%]	d.r.
1	*p*-TsOH	5.0	toluene	25	24	40–60[a]	2.5:1[a]
2	PPTS	3.0	toluene	100	1	39	3.3:1
3	H_3PO_4	3.0	THF	25	24	38	5.5:1
4	(*R*)-binol·HPO_4	1.0	$CHCl_3$	100[b]	1	63	3.9:1
5	(*S*)-binol·HPO_4	1.0	$CHCl_3$	100[b]	1	62	3.9:1
6	*rac*-binol·HPO_4	1.0	$CHCl_3$	100[b]	1	55	3.0:1
7	(*R*)-binol·HPO_4	1.0	$CHCl_3$	25	24	32	>10:1
8	(*R*)-binol·HPO_4	1.0	DMSO	25	24	33	1.8:1
9	(*R*)-binol·HPO_4	1.0	$CHCl_3$/MeOH (5:1)	25	24	41	5.0:1
10	(*R*)-binol·HPO_4	0.7	$CHCl_3$	100[b]	1	55	4.5:1
11	(*S*)-binol·HPO_4	0.7	$CHCl_3$	100[b]	1	54	4.6:1
12	*rac*-binol·HPO_4	0.7	$CHCl_3$	100[b]	1	50	3.1:1
13	(*R*)-**21**	1.0	$CHCl_3$	100[b]	1	56	18.4:1
14	(*S*)-**21**	1.0	$CHCl_3$	100[b]	1	56	18.9:1
15	*rac*-**21**	1.0	$CHCl_3$	100[b]	1	59	13.6:1
16	(*R*)-CSA	2.0	$CHCl_3$	50	2	54	4.0:1
17	(*S*)-CSA	2.0	$CHCl_3$	50	2	56	4.1:1
18	*rac*-CSA	2.0	$CHCl_3$	50	2	53	4.0:1

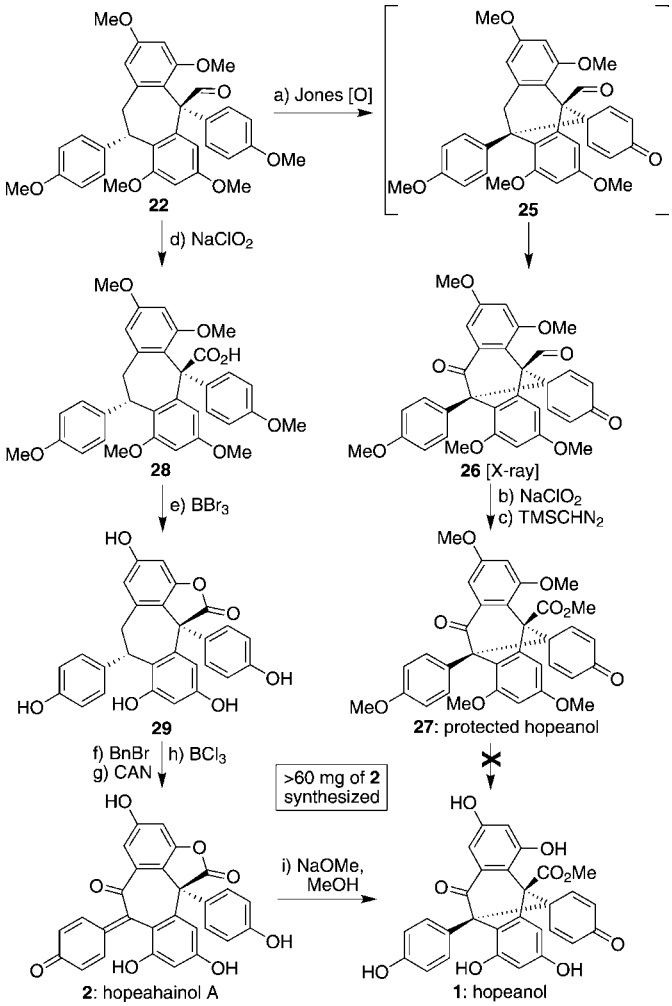
[a] Both yield and d.r. proved highly variable between runs; d.r. as high as 4:1 for **22**/**23** were observed, but 2.5:1 was more common, especially on a large scale.
[b] Under microwave irradiation.

collated in Table 1, entries 1–3. However, the most important and consistent observation in all experiments was that diol diastereomer **19** transformed into **22** quickly and with high diastereoselectivity (typically greater than 10:1), while **20** reacted much more slowly, provided more side products, and required increased reaction temperatures for any conversion (leading to **22** and **23**).[21]

As such, the goal for optimization became finding an acid source with a suitable pK_a value capable not only of rearranging **19** smoothly, but also improving the throughput of **20**. Our first significant advance based on this analysis occurred when a mixture of both **19** and **20** was stirred with one equivalent of (*R*)-binol·HPO_4[22] in $CHCl_3$ at 100 °C under microwave irradiation for 1 h. These conditions led to pinacol-rearranged products **22** and **23** in 63 % yield and 3.9:1 diastereocontrol in favor of **22** (Table 1, entry 4) alongside varying amounts of epoxide **24** (ca. 10–15 %).[23] Other solvents and conditions with this promoter afforded decreased selectivity and/or yield (Table 1, entries 7–9) for **22**. Interestingly, while use of the opposite enantiomer of promoter [(*S*)-binol·HPO_4] under these conditions afforded nearly identical results, its racemic form provided inferior stereoselection (Table 1, entries 5 and 6).[24] The same phenomenon was also observed when decreased quantities of phosphoric acid were used, though these cases afforded improved diastereoselectivity (4.5:1) at the price of yield (Table 1, entries 10–12). It was also observed when the promoter size was changed to that of vapol·HPO_4 (**21**, Table 1, entries 13–15).[25] In these cases, high diastereoselection (greater than 18:1) and similar throughput efficiency (56 % yield of **22** and **23**; trace **24**) was achieved when a single enantiomer was used. At present, it is too preliminary to provide a rationale for these unexpected outcomes between the use of racemic or single enantiomer forms of these promoters other than to state that it is a reproducible result over several runs. We do note, however, that this effect may be specific to materials of the binol and vapol scaffolds in that both chiral and racemic camphorsulfonic acid (CSA) gave nearly identical results (Table 1, entries 16–18).[26] Current work is directed at understanding the parameters of this event more fully, especially determining whether chiral acids have value in other pinacol rearrangements where incongruities exist in diastereomer reactivity and stereoselection. What we believe we can state is that, to the best of our knowledge, this event constitutes the first use of a chiral Brønsted acid for this rearrangement in a total synthesis.

With this key quaternary carbon center forged with good efficiency, our next goal was to effect the remaining oxidations needed to access the natural products. These steps had to install the missing ketone located at the C8a-position, convert the hindered aldehyde into a carboxylic acid, and generate the C–C bond leading to the dearomatized *p*-quinone ring essential to hopeanol (**1**). After an exhaustive screen of oxidants, including 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ), hypervalent iodine, and $Pd(OAc)_2/H_2O_2$,[27] we discovered that the Jones reagent could uniquely accomplish two of these tasks when it was added to an acetone solution of **22** at 0 °C and stirred for 30 min (Scheme 3). This step leading to **26** proceeded in 27 % overall yield, with only the drawn diastereomer of **22** reacting productively.[28] Equally intriguing, this event appears to proceed by the initial formation of **25**, as a trace amount of this material was obtained when insufficient Jones reagent was available to drive the reaction to completion; this material (that is, **25**) was quickly converted into **26** following re-exposure to the Jones reagent. Surprisingly, the aldehyde within **22** was not oxidized in this step; thus, a different oxidant ($NaClO_2$) proved necessary. Then, following treatment with $TMSCHN_2$, protected hopeanol (**27**) was obtained in 75 % yield (4.3 % overall from **15**). Despite much effort, however, this material could not be deprotected,[29] including use of the conditions of Nicolaou et al.[5]

As such, efforts were made to deprotect the phenolic methyl ether groups earlier in the sequence, such as at the stage of aldehyde **26** and intermediate **22**. Unfortunately, in both cases (as well as many others not explicitly described here), these attempts consistently led to rearrangement reactions and/or decomposition, one of which is denoted in the Supporting Information section. Pleasingly, when carboxylic acid **28** was treated with BBr_3 in CH_2Cl_2, methyl ether cleavage was attended by lactone formation to afford **29**; a small amount of decarboxylated material was also observed.



Scheme 3. Completion of hopeanol (**1**) and hopeahainol A (**2**). a) Jones reagent (20 equiv), acetone, 0 °C, 30 min, 27 %. b) Resorcinol (15 equiv), $NaH_2PO_4{\cdot}H_2O$ (10 equiv), $NaClO_2$ (5 equiv), THF/*t*BuOH/H_2O (1:1:2), 25 °C, 3 days, 70 %. c) $TMSCHN_2$ (5 equiv), THF/MeOH (4:1), 0 °C, 15 min, 98 %. d) Resorcinol (15 equiv), $NaH_2PO_4{\cdot}H_2O$ (10 equiv), $NaClO_2$ (5 equiv), THF/*t*BuOH/H_2O (1:1:2), 25 °C, 3 days. e) BBr_3 (18 equiv), CH_2Cl_2, −78→0 °C, 3 h. f) BnBr (20 equiv), K_2CO_3 (20 equiv), nBu_4NI (1.0 equiv), acetone, 25 °C, 12 h, 29 % over three steps. g) CAN (8.0 equiv), DMSO, 25 °C, 12 h, 65–89 %. h) BCl_3 (12 equiv), CH_2Cl_2, −78 °C, 15 min, 75 %. i) NaOMe (1.2 equiv), MeOH, 25 °C, 4 days, 69 %.

Although all attempts at oxidizing this material directly to **1** or **2** failed, its benzyl ether analogue could be oxidized with ceric ammonium nitrate (CAN) to afford protected hopeahainol A in 65–89 % yield, depending on the scale. No other oxidant succeeded. Finally, deprotection with BCl_3 then delivered the natural product (**2**) in 75 % yield, a portion of which was converted into hopeanol (**1**) following the exact conditions developed by Nicolaou et al.[5] In total, the route to hopeahainol A (**2**) is 14 steps long, and as one reflection of its overall efficiency (4.0 % overall from **15**), we have prepared over 60 mg of it along with 180 mg of its protected precursor to date.

In conclusion, we have accomplished an efficient total synthesis of both hopeanol (**1**) and hopeahainol (**2**) from our key precursor for the controlled preparation of the resveratrol family (that is, **7**) through a pathway that traces both of these structures to a more common manifold within this fascinating oligomer family. Critical steps involved a pinacol rearrangement empowered by a chiral phosphoric acid and multistage, substrate-specific oxidation processes. Current efforts are directed towards developing asymmetric syntheses of these materials and probing their chemical biology.

Received: November 2, 2011
Revised: February 15, 2012
Published online: March 16, 2012

Keywords: hopeahainol A · hopeanol · natural products · resveratrol · total synthesis

[1] For reviews, see: a) S. Quideau, D. Deffieux, C. Douat-Casassus, L. Pouységu, *Angew. Chem.* **2011**, *123*, 610–646; *Angew. Chem. Int. Ed.* **2011**, *50*, 586–621; b) S. A. Snyder in *Biomimetic Organic Synthesis* (Ed.: E. Poupon, B. Nay), Wiley-VCH, Weinheim, **2011**, pp. 695–721.

[2] For recent papers concerning the bioactivity of resveratrol itself, see: a) M. Jang, L. Cai, G. O. Udeani, K. V. Slowing, C. F. Thomas, C. W. W. Beecher, H. H. S. Fong, N. R. Farnsworth, A. D. Kinghorn, R. G. Mehta, R. C. Moon, J. M. Pezzuto, *Science* **1997**, *275*, 218–220; b) K. T. Howitz, K. J. Bitterman, H. Y. Cohen, D. W. Lamming, S. Lavu, J. G. Wood, R. E. Zipkin, P. Chung, A. Kisielewski, L.-L. Zhang, B. Scherer, D. A. Sinclair, *Nature* **2003**, *425*, 191–196; c) J. G. Wood, B. Rogina, S. Lavu, K. Howitz, S. L. Helfand, M. Tatar, D. Sinclair, *Nature* **2004**, *430*, 686–689, and references therein.

[3] For selected, recent examples of synthetic efforts to prepare resveratrol oligomers, see: a) W. Li, H. Li, Y. Li, Z. Hou, *Angew. Chem.* **2006**, *118*, 7771–7773; *Angew. Chem. Int. Ed.* **2006**, *45*, 7609–7611; b) J. Zhu, C. Zhong, H.-F. Lu, G.-Y. Li, X. Sun, *Synlett* **2008**, 458–462; c) S. S. Velu, I. Buniyamin, L. K. Ching, F. Feroz, I. Noorbatcha, L. C. Gee, K. Awang, I. A. Wahab, J.-F. F. Weber, *Chem. Eur. J.* **2008**, *14*, 11376–11384; d) J. L. Jeffrey, R. Sarpong, *Tetrahedron Lett.* **2009**, *50*, 1969–1972; e) I. Kim, J. Choi, *Org. Biomol. Chem.* **2009**, *7*, 2788–2795; f) J. L. Jeffrey, R. Sarpong, *Org. Lett.* **2009**, *11*, 5450–5453; g) W. Li, Y. Luo, H. Li, P. Zang, X. Han, *Synthesis* **2010**, 3822–3826; h) C. Zhong, J. Zhu, J. Chang, X. Sun, *Tetrahedron Lett.* **2011**, *52*, 2815–2817; i) Y. Yang, D. Philips, S. Pan, *J. Org. Chem.* **2011**, *76*, 1902–1905.

[4] a) H. M. Ge, C. Xu, X. T. Wang, B. Huang, R. X. Tan, *Eur. J. Org. Chem.* **2006**, 5551–5554; b) H. M. Ge, C. H. Zhu, D. H. Shi, L. D. Zhang, D. Q. Xie, J. Yang, S. W. Ng, R. X. Tan, *Chem. Eur. J.* **2008**, *14*, 376–381.

[5] a) K. C. Nicolaou, T. R. Wu, Q. Kang, D. Y.-K. Chen, *Angew. Chem.* **2009**, *121*, 3492–3495; *Angew. Chem. Int. Ed.* **2009**, *48*, 3440–3443; b) K. C. Nicolaou, Q. Kang, T. R. Wu, C. S. Lim, D. Y.-K. Chen, *J. Am. Chem. Soc.* **2010**, *132*, 7540–7548.

[6] For a recent review on cascade reactions, see: K. C. Nicolaou, D. J. Edmonds, P. G. Bulger, *Angew. Chem.* **2006**, *118*, 7292–7344; *Angew. Chem. Int. Ed.* **2006**, *45*, 7134–7186.

[7] For examples using the pinacol reaction to forge quarternary carbon centers in natural product synthesis, see: a) J. Li, S. Jeong, L. Esser, P. G. Harran, *Angew. Chem.* **2001**, *113*, 4901–4906; *Angew. Chem. Int. Ed.* **2001**, *40*, 4765–4769; b) G. R. Pettit, J. W. Lippert, D. L. Herald, *J. Org. Chem.* **2000**, *65*, 7438–7444.

[8] a) S. A. Snyder, A. L. Zografos, Y. Lin, *Angew. Chem.* **2007**, *119*, 8334–8339; *Angew. Chem. Int. Ed.* **2007**, *46*, 8186–8191; b) S. A.

Snyder, S. P. Breazzano, A. G. Ross, Y. Lin, A. L. Zografos, *J. Am. Chem. Soc.* **2009**, *131*, 1753 – 1765.

[9] For our work towards other resveratrol-based structures, see: a) S. A. Snyder, A. Gollner, M. I. Chiriac, *Nature* **2011**, *474*, 461 – 466; b) S. A. Snyder, N. E. Wright, J. J. Pflueger, S. P. Breazzano, *Angew. Chem.* **2011**, *123*, 8788 – 8792; *Angew. Chem. Int. Ed.* **2011**, *50*, 8629 – 8633; c) S. A. Snyder, Z. G. Brill, *Org. Lett.* **2011**, *13*, 5524 – 5527.

[10] For reviews, see: a) N. Z. Burns, P. S. Baran, R. W. Hoffmann, *Angew. Chem.* **2009**, *121*, 2896 – 2910; *Angew. Chem. Int. Ed.* **2009**, *48*, 2854 – 2867; b) T. Newhouse, P. S. Baran, R. W. Hoffmann, *Chem. Soc. Rev.* **2009**, *38*, 3010 – 3021.

[11] Although these exact precursors are not known isolates, there are natural products where the putative double bond of **8** – **10** is reduced. For an example, see: K. Baba, Y. Tabata, K. Maeda, M. Doi, M. Kozawa, *Chem. Pharm. Bull.* **1986**, *34*, 4418 – 4421.

[12] For the isolation of these natural products, see: a) Y. Oshima, Y. Ueno, H. Hikino, L. L. Yang, K. Y. Yen, *Tetrahedron* **1990**, *46*, 5121 – 5126; b) T. Ito, T. Tanaka, M. Iinuma, K. Nakaya, Y. Takahashi, R. Sawa, J. Murata, D. Darneadi, *J. Nat. Prod.* **2004**, *67*, 932 – 937; c) J.-F. F. Weber, I. A. Wahab, A. Marzuki, N. F. Thomas, A. A. Kadir, A. H. A. Hadi, K. Awang, A. A. Latiff, P. Richomme, J. Delaunay, *Tetrahedron Lett.* **2001**, *42*, 4895 – 4897; d) H. M. Ge, W.-H. Yang, J. Zhang, R.-X. Tan, *J. Agric. Food Chem.* **2009**, *57*, 5756 – 5761.

[13] If this proposal is accurate, it would mean that these dimers result from the reorganization of bonds within a dimeric framework, rather than a union of resveratrol monomers as originally proposed (Ref. [4]).

[14] This route is shorter than our previously published procedures (Ref. [8]) because of a change in commercial starting material. See the Supporting Information for full details.

[15] The drawn orientation of **15** is based on a crystal structure of a closely related compound (see the Supporting Information) indicating the pseudoaxial orientation of the pendant aryl ring on the seven-membered ring as well as the positioning of the ketone relative to the neighboring aryl substituents.

[16] E. J. Corey, M. Chaykovsky, *J. Am. Chem. Soc.* **1965**, *87*, 1353 – 1364.

[17] The intermediate acetate was never observed and likely hydrolyzes on work-up; in practice, the AcOH reaction and oxidation were performed as a single step without intervening chromatography.

[18] Extensive efforts to characterize **19** and **20** by crystallization failed; the current assignment is based on nOe experiments as noted in the Supporting Information.

[19] As evidence for the existence of **17**, the pendant aryl ring could form a disubstituted quinone methide in 11% yield (see the Supporting Information).

[20] This outcome suggests that semipinacol processes from **24** are not operative.

[21] The basis for the differential diastereoselectivity of **19** and **20** in the pinacol rearrangement is not obvious based on simple molecular models, though it is clear based on experimental observations that diol **19** has a lower energy barrier for the conversion.

[22] For the pioneering use of such species in related rearrangements, see: T. Liang, Z. Zhang, J. C. Antilla, *Angew. Chem.* **2010**, *122*, 9928 – 9930; *Angew. Chem. Int. Ed.* **2010**, *49*, 9734 – 9736.

[23] Neither **22** nor **23** were generated with optical activity.

[24] Racemic binol·HPO_4, prepared from racemic binol or a mixture of both (*S*)- and (*R*)-forms of the reagent, gave the same results.

[25] A. A. Desai, L. Huang, W. D. Wulff, G. B. Rowland, J. C. Antilla, *Synthesis* **2010**, 2106 – 2109.

[26] Globally, we believe that the overall success of these processes is related to the pK_a value of the acids used, though exact values for their acidity across the range of solvents used in this study are not known.

[27] a) J.-Q. Yu, E. J. Corey, *Org. Lett.* **2002**, *4*, 2727 – 2730; b) J.-Q. Yu, E. J. Corey, *J. Am. Chem. Soc.* **2003**, *125*, 3232 – 3233.

[28] The chemospecificity of this oxidation process is intriguing. For related examples of high reagent/substrate specificity, see: S. A. Snyder, T. C. Sherwood, A. G. Ross, *Angew. Chem.* **2010**, *122*, 5272 – 5276; *Angew. Chem. Int. Ed.* **2010**, *49*, 5146 – 5150.

[29] Other deprotections include: a) S. J. O'Malley, K. L. Tan, A. Watzke, R. G. Bergman, J. A. Ellman, *J. Am. Chem. Soc.* **2005**, *127*, 13496 – 13497; b) K. C. Nicolaou, M. Takayanagi, N. F. Jain, S. Natarajan, A. E. Koumbis, T. Bando, J. M. Ramanjulu, *Angew. Chem.* **1998**, *110*, 2881 – 2883; *Angew. Chem. Int. Ed.* **1998**, *37*, 2717 – 2719.

